



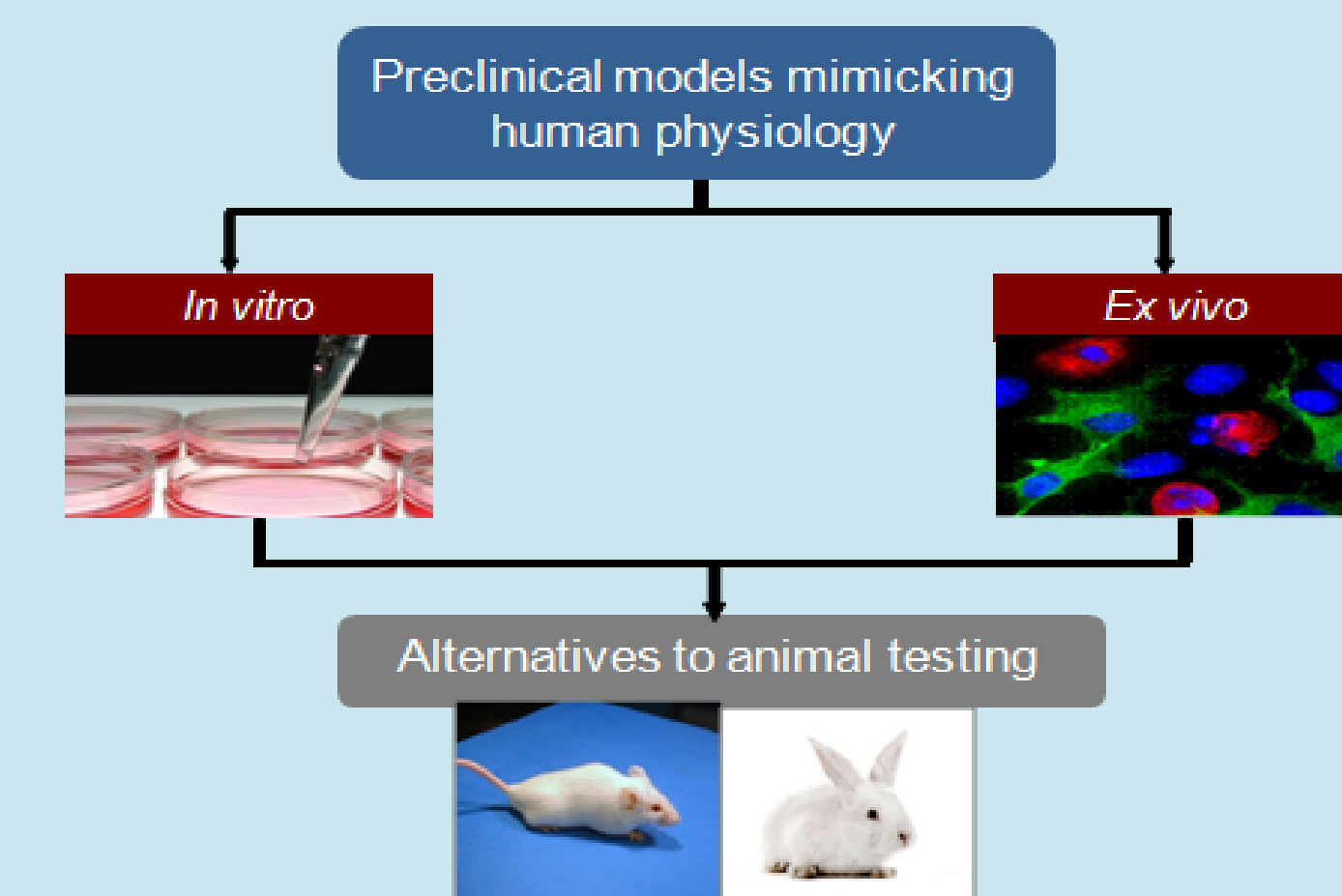
ABSTRACT

Screening of biological activities of new therapeutic compounds using in vitro assays provides a fast and reliable measure of efficacy and safety. These cell-based assays not only potentially accelerate the research of new therapeutic agents, but may also serve as alternatives to equivalent animal tests in vivo by reducing the number of animals and severity of procedures. At Dabur Research Foundation(DRF), we have developed a repertoire of in vitro models to assess activities of test agents in multiple therapeutic areas. Cytotoxic profile of anticancer compounds is evaluated in a panel of human and murine cancer cell lines with safety assessment in normal cell lines. Murine and human bone marrow cells derived Colony Forming Unit (CFU-GM) assays are employed to predict hematotoxic side-effects of anticancer drugs, which is European Commission for Validation of Alternative Methods (ECVAM) approved method to determine human MTD for neutropenia. Specialized cell-based screening models have been developed in the area of inflammation, such as Dendritic cells for systemic inflammation, keratinocytes/monocytes for psoriasis, lung/nasal epithelium cells for airway inflammation and allergy, fibroblasts for dermal inflammation and intestinal/colon epithelium cells for gastrointestinal-inflammation. Sophisticated in vitro models based on keratinocytes and sebocytes are available to screen anti-acne properties of cosmetic/dermatological products. Skin health parameters such as anti-aging and anti-wrinkling potential are explored using skin-fibroblast cell lines. These in vitro assay systems contribute towards understanding of complex biological action of new compounds at cellular level. Shortlisting of hit/active compounds by in-vitro screening obviates the need of large number of testing animals while meeting 3R's principle of "Reduce, Refine and Replace".

INTRODUCTION

Animal models have been widely employed to assess pharmacological activity of new compounds. To predict toxicity, corrosivity, and other safety variables in addition to effectiveness of a new product for humans, traditional testing of new drugs involves the use of animals on a large scale. Registration, Evaluation and Authorization of Chemicals (REACH; regulation 1907/2006) is in effect in the European Union (EU) and safety information must now be provided on all chemicals that are either sold, manufactured or imported into the EU. Furthermore, an animal testing ban has been effective on chemicals to be used in cosmetics in EU from March 2009. Realizing the importance in drug discovery, many alternative methods for animal testing are being developed, which have shown to lead to safer and more effective products for humans. Alternative testing methods have many advantages over traditional animal tests-including being more humane, reliable, accurate, more cost-effective, fast, practical and expedient. We have developed a huge spectrum of in vitro/ex vivo cell-based models in varied therapeutic applications such as cancer, hematotoxicity, immunomodulation, inflammation, dermatology and skin-care. These models profoundly increase the understanding of biological activities of test compounds and may serve as in vitro alternatives to similar tests conducted in animals.

- Reduction - To minimize number of animals used
Refinement - To minimize suffering and distress
Replacement - To avoid the use of living animals



IMMUNITY & INFLAMMATION

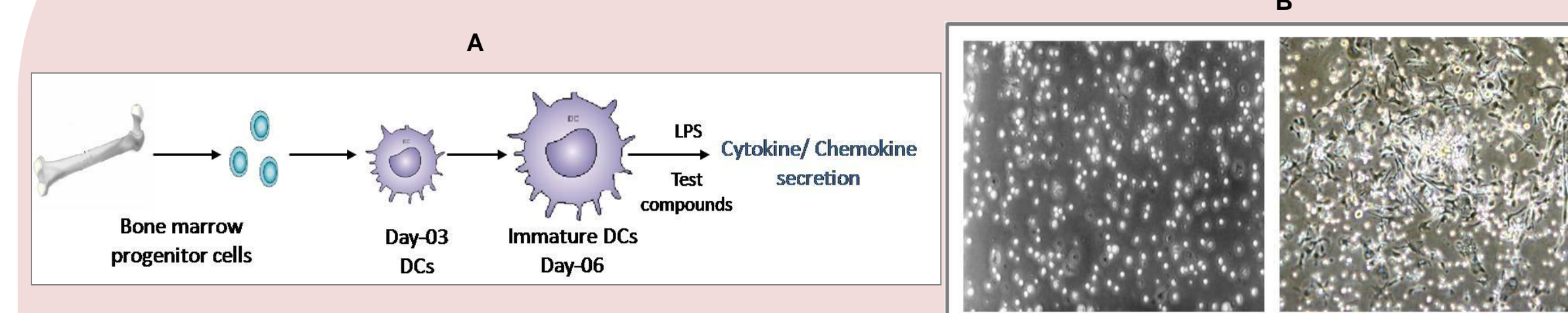


Figure 1 - Dendritic Cells (DC) assay (A) Generation of primary DC cultures from murine bone marrow (B) Morphology of day-3 and day-6 DCs

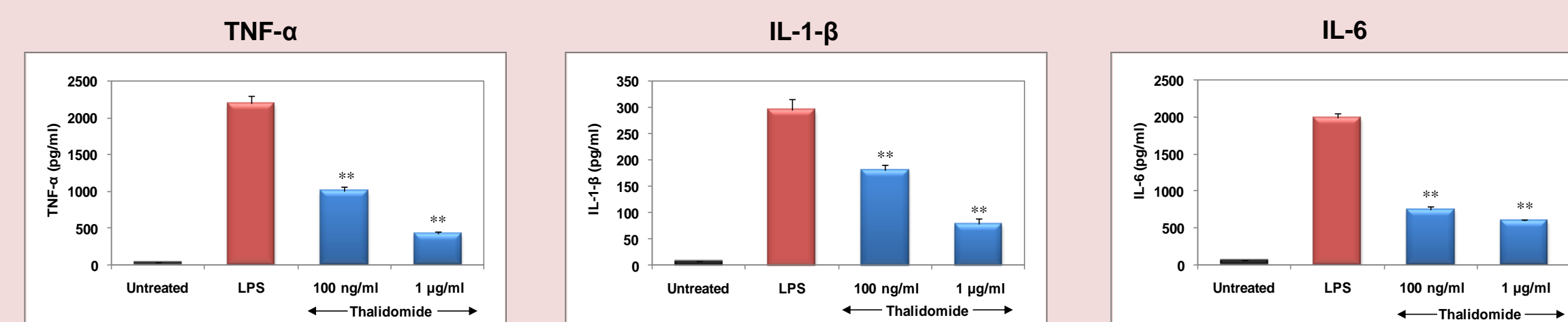


Figure 2 - Anti-inflammatory effect of Thalidomide in DCs based model

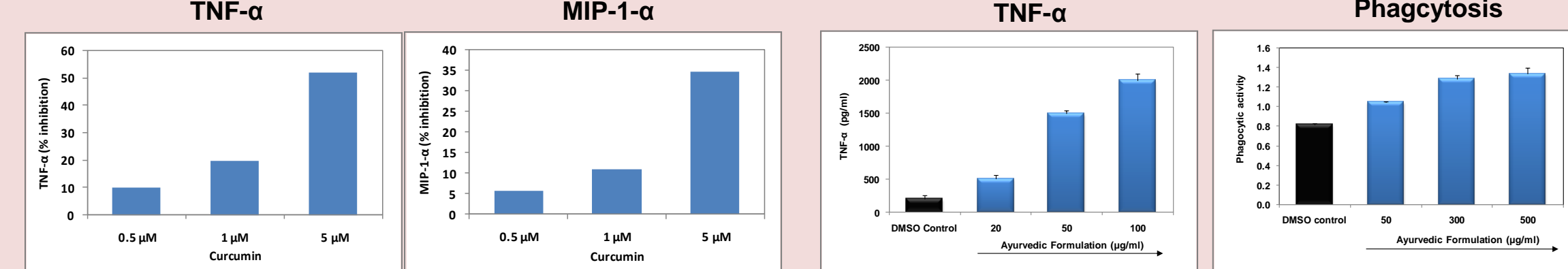


Figure 3 - Anti-inflammatory effect of Thalidomide in primary murine splenocytes

Figure 4 - Immunostimulatory effect of an ayurvedic formulation in DCs and macrophages (RAW264.7)

- Comprehensive range of in vitro & ex vivo models for identification of immunomodulatory and anti-inflammatory potential
Cytokine and chemokine profiling in murine bone marrow derived DCs
Phagocytosis in murine macrophages (RAW264.7)
Anti-inflammatory effect by inhibition of LPS induced cytokine/chemokine in DCs and splenocytes

ANTICANCER ACTIVITY

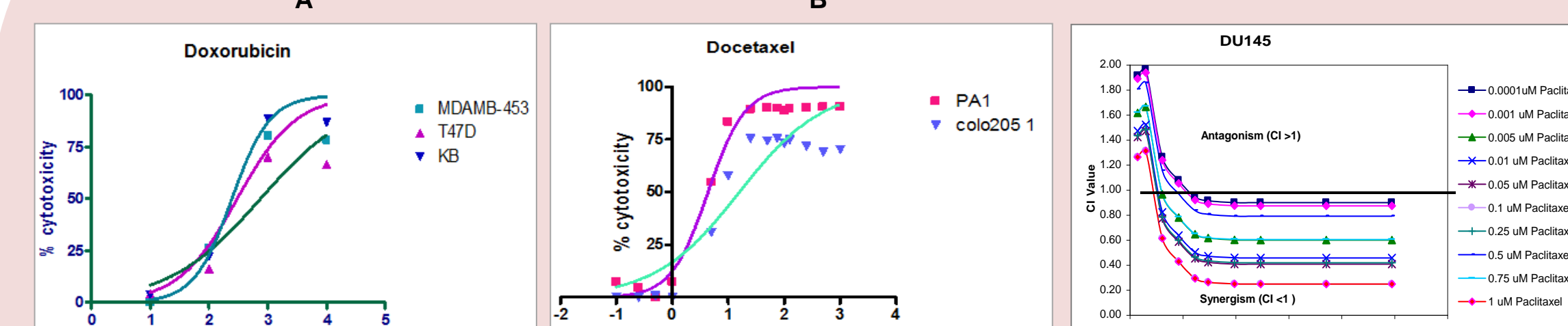


Figure 6 - Cytotoxic profile of (A) Doxorubicin and (B) Docetaxel in human cancer cell lines

Figure 7 - Synergistic cytotoxic profile of Paclitaxel and Quercetin in human prostate cancer cell line (DU145)

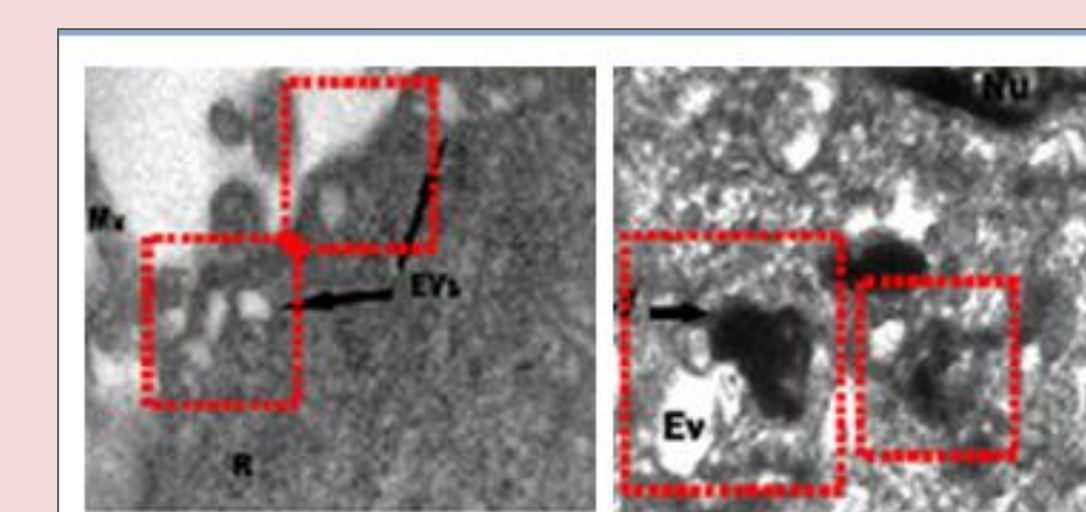


Figure 8 - Transmission electron micrographs of human breast cancer (MDA-MB-453) and (B) PA1 (human ovarian cancer) cells treated with nanoparticle formulation (60kx)

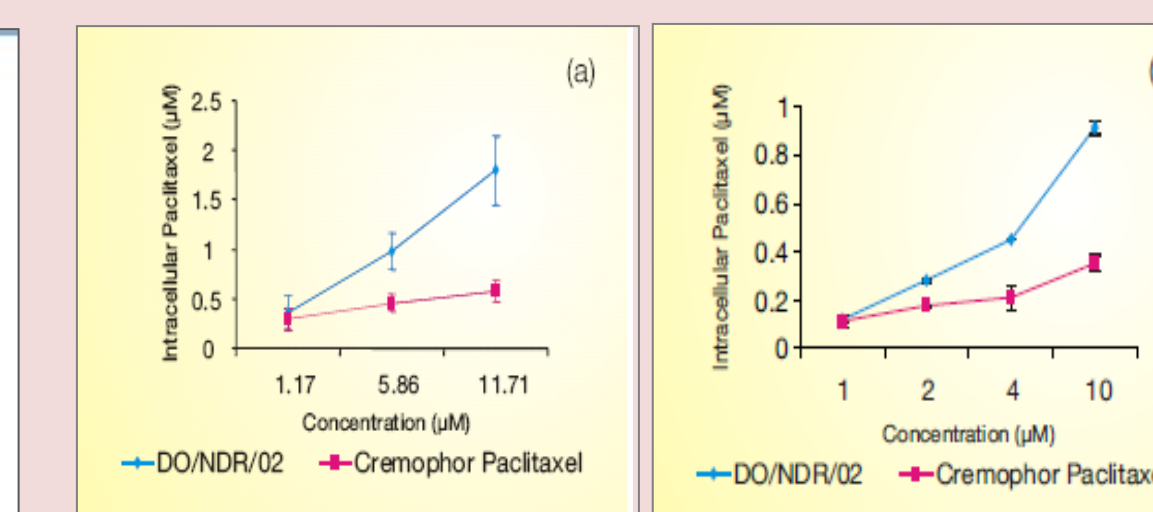


Figure 9 - Intracellular accumulation of Paclitaxel in (A) MDA-MB-453 and (B) PA1 (human ovarian cancer) cells treated with nanoparticle formulation

- Cytotoxicity screening in human and murine cancerous cell lines representing 15 - 20 different cancer types (ATCC/ECACC)
Safety index in normal cell lines
Synergistic/additive/antagonistic anticancer activity by combination index method
Efficacy of targeted drugs
Mechanism of drug uptake and subcellular distribution by Transmission electron Microscopy (TEM) and HPLC method
Angiogenesis in endothelial cell based assay

DERMATOLOGY

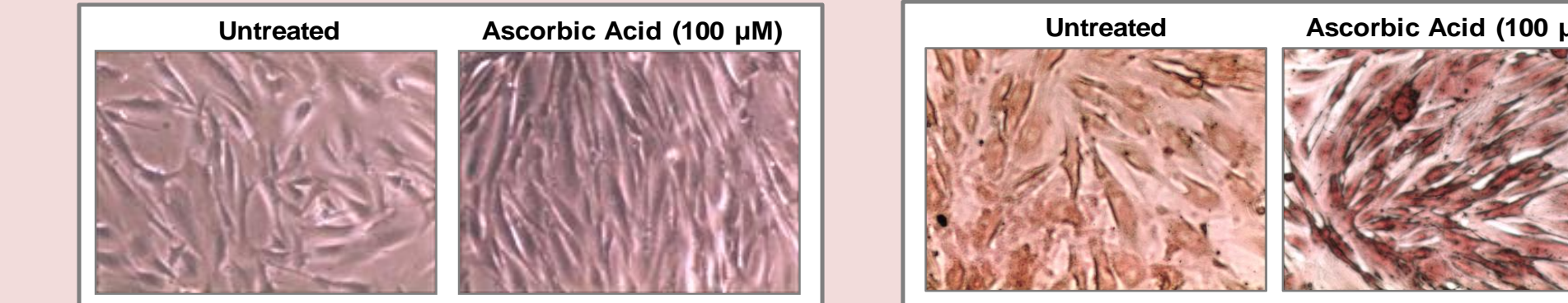


Figure 12 - Skin-health promoting effects of Ascorbic acid (100 µM) on (A) Morphology (B) Collagen synthesis in human fibroblast cell line (HFF-1)

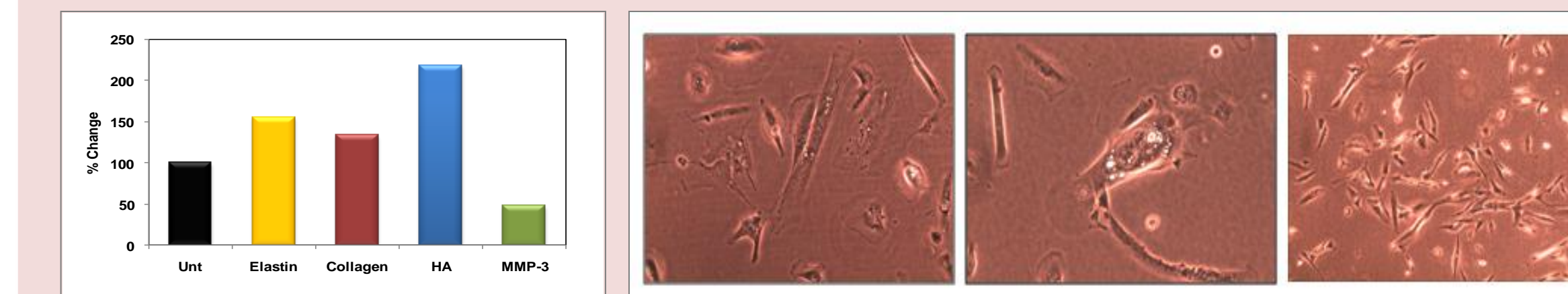


Figure 13 - Effect of Ascorbic acid on skin-health parameters in HFF-1 cells

Figure 14 - Morphology of cultured human sebocytes

- In vitro screens to assess cosmetic/dermatological products

Table with 4 columns: No., Indication, Model, End points. Rows include Skin-health, Anti-aging/Anti-wrinkling, Skin-lightening/Vitiligo, Dermatitis, and Anti-acne.

RESPIRATORY INFLAMMATION/ALLERGY

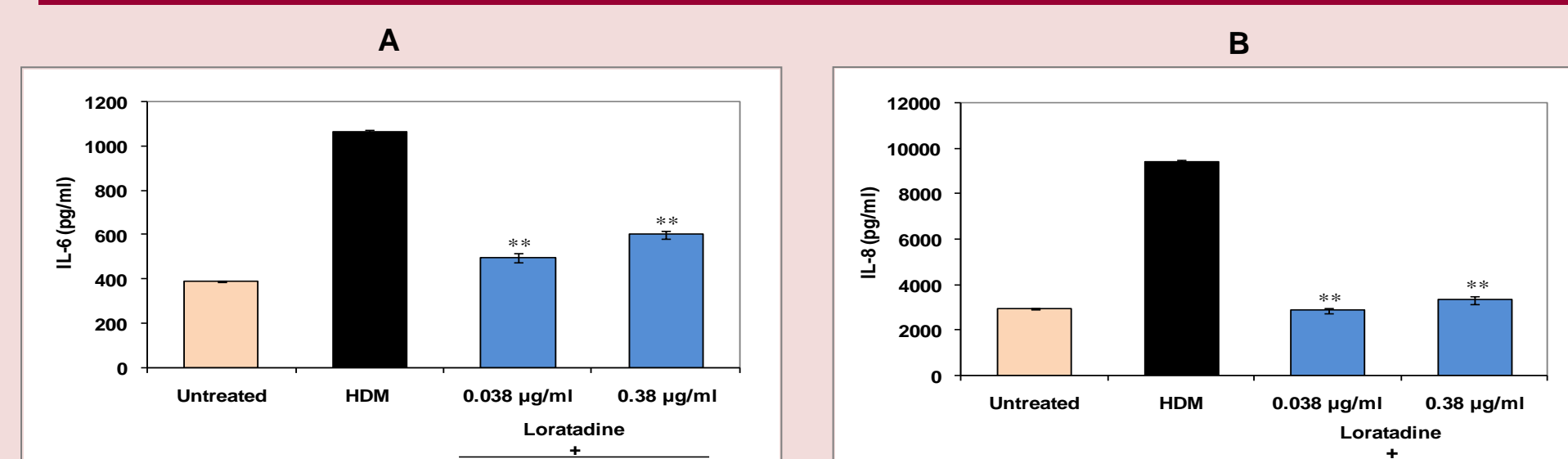


Figure 5 - Inhibitory effects of Loratadine on HDM-induced cytokine secretion (A) IL-6 and (B) IL-8 in human alveolar epithelium cell line (A-549)

- Downregulation of inflammatory markers in cell lines representing airway epithelium:

Table with 4 columns: No., Indication, Test system, Model, End points. Rows include Anti-allergic/Pulmonary inflammation, Rhinitis, Pulmonary inflammation/allergy, and Pulmonary inflammation/allergy.

HEMATOTOXICITY

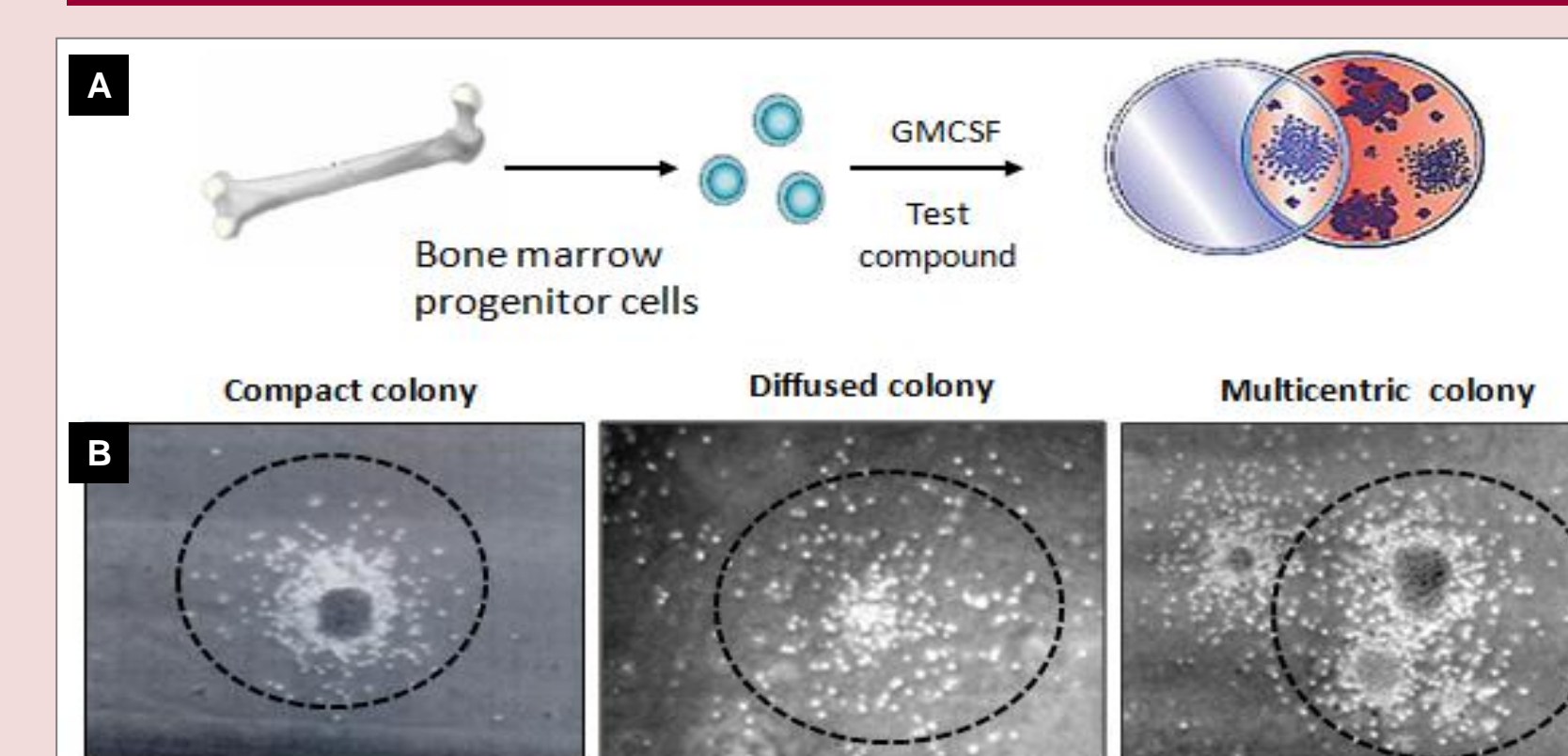
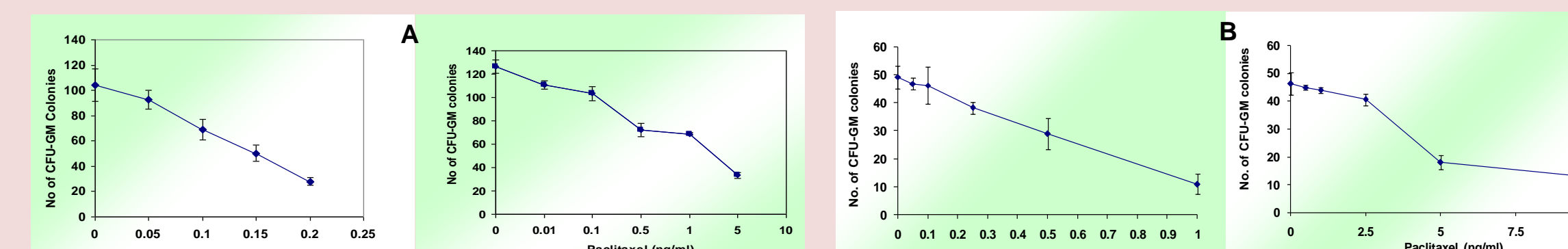


Figure 10 - CFU-GM assays (A) Assay design (B) Characteristic morphology of CFU-GM colonies



- In vitro Colony Forming Unit (CFU) assays serve as predictive models for chemotherapy induced hematotoxicity

Table with 3 columns: CFU in hematopoiesis, Growth factors, Type of prediction. Rows include CFU-GM (Granulocyte-macrophage), CFU-E, BFU-E (Erythrocytes), and CFU-Meg (Platelets).

- For anticancer drugs with neutropenia as dose-limiting toxicity, CFU-GM assay is approved by ECVAM Ispra, Italy (24th meeting, March 2006) to predict acute neutropenia in humans

Predicted human MTD = Actual murine LD10 x IC human CFU-GM assay / IC murine CFU-GM assay

PSORIASIS

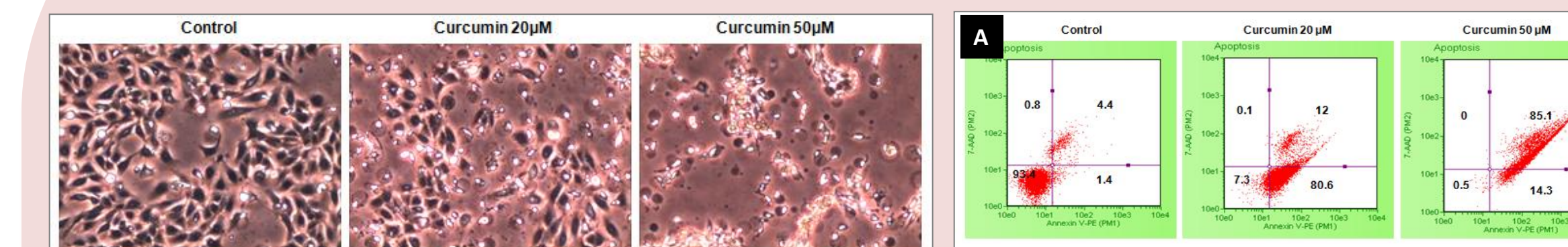


Figure 15 - Effect of Curcumin on morphology of human keratinocyte cell line (HaCaT) after 24 h

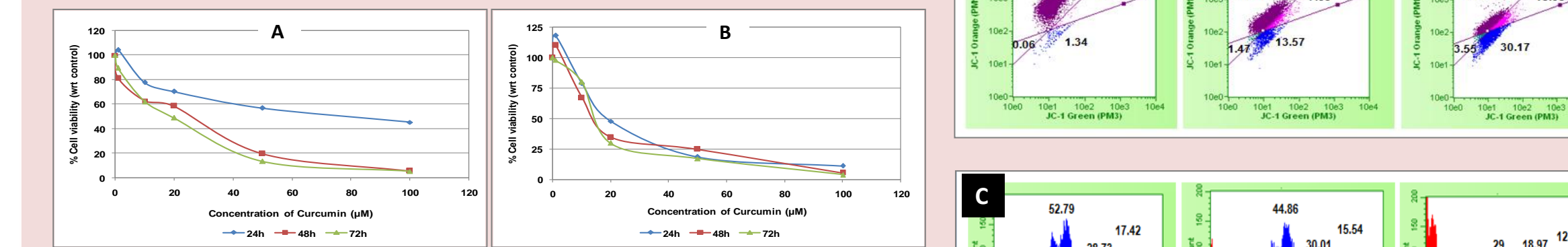


Figure 16 - Anti-proliferative effects of Curcumin in (A) HaCaT and (B) human monocytic cell line (THP-1) after 24, 48 and 72 h

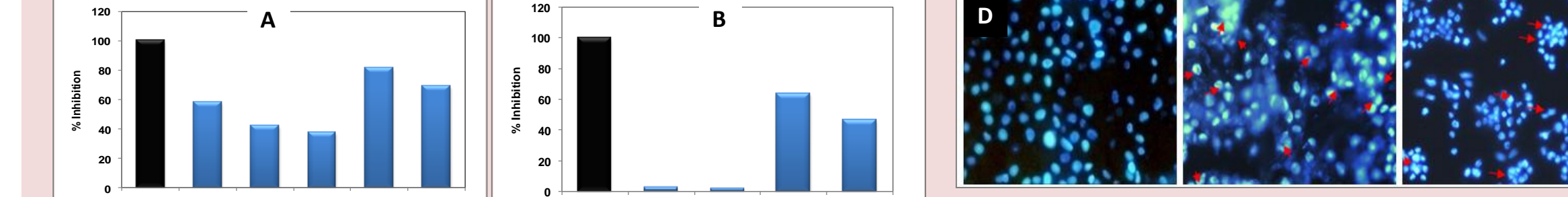


Figure 17- Anti-inflammatory effects of Curcumin (20µM) in (A) HaCaT and (B) THP-1 cells after 24 h against TNF-alpha and LPS stimulation respectively

Figure 18- Induction of apoptosis by Curcumin in HaCaT cells (A) Annexin-V (B) Mitochondrial potential (C) Cell cycle (D) DNA fragmentation after 24 h

- HaCaT (human keratinocytes) and THP-1 (human monocytes) cells based model to evaluate anti-psoriatic potential
Anti-proliferative effects - Time and dose kinetics
Anti-inflammatory activity
Inhibition of cytokines in HaCaT cells against TNF-alpha stimulation
Inhibition of cytokines in THP-1 cells against LPS stimulation
Pro-apoptotic effect
Increase in Annexin-V+ cells, loss of mitochondrial potential, increase in sub(G0/G1) population and DNA fragmentation